

PII: S0959-8049(97)00096-8

Original Paper

p53 Expression is of Independent Predictive Value in Lymph Node-negative Breast Carcinoma

M. Fresno, R. Molina, M.J. Pérez del Río, S. Alvarez, J.M. Díaz-Iglesias, I. García and A. Herrero

Department of Pathology, Covadonga University Hospital, University of Oviedo, Celestino Villamil S/N 33006 Oviedo, Spain

The aim of this study was to evaluate p53 expression, determined by immunohistochemistry, in 151 infiltrating ductal breast carcinomas with negative axillary lymph nodes, and to determine whether p53 can be considered as an independent prognostic value for overall and disease-free survival. A monoclonal antibody (DO-7) that reacts with an epitope on the N terminal portion of the human protein p53 was used to detect p53 in paraffin-embedded sections, utilising a standard avidin-biotin-peroxidase complex (ABC) technique with a microwave oven antigen retrieval. Overexpression of p53 (more than 50% of stained cells) was found in 45 cases (30%). Forty-five cases were negative and occasionally or moderately stained cells were present in 61 cases. p53 protein overexpression was significantly associated with high histological grade and tumour necrosis, high MIB-1 value (MIB-1>30%) and negative oestrogen receptor status. Univariate analysis (log-rank) showed a shorter overall survival (P = 0.003) in patients with high tumour p53 positivity. This statistical significance was also seen on multivariate analysis (Cox's logistic regression, P = 0.004). p53 protein overexpression is an independent prognostic marker in node-negative breast carcinoma for overall survival and should be used with other prognostic factors. © 1997 Published by Elsevier Science Ltd.

Key words: p53, immunohistochemistry, oncoprotein, breast carcinoma

Eur J Cancer, Vol. 33, No. 8, pp. 1268-1274, 1997

INTRODUCTION

AXILLARY LYMPH node status is currently the most important prognostic factor for breast cancer. However, 20–30% of women with node-negative tumours experience recurrence within 5 years. Thus, the search for additional tumour markers with prognostic value has increased considerably in the last few years, especially in node-negative tumours [1].

The discovery of tumour suppressor genes has provided a new approach to our understanding of tumour cell biology. One of the best known tumour suppressor genes is the *TP53* gene [2–4]. Point mutations or deletion of the wild type gene have been suggested as a key event in the development of malignancy [5, 6]. The wild type p53 protein is present in the nuclei of all mammalian cells where it appears

to be involved in the regulation of cell proliferation, acting as a suppressor of cell growth [7-9]. In the normal cell, the concentration of the wild type p53 protein is generally below the detection level for immunohistochemical procedures. However, tumours with the mutant form of the TP53 gene may express high levels of p53 protein with a significantly prolonged half-life, which can reach the threshold of immunoreactivity [10]. Although, in some studies, a correlation between overexpression of p53 protein and mutation of the TP53 gene has been found, others have found discrepancies [11, 12]. It is believed that, in some cases, wild type p53 can be overexpressed or stabilised by nonmutational factors. Overexpression of p53 could interfere with cell-cycle regulatory functions of the wild type protein. This mutated TP53 acts as an oncogene [7, 13, 14]. Overexpression of p53 in breast [15-19], ovarian [20, 21], lung [10, 22], bladder [23-25] and colon carcinomas [26, 27] has been associated with aggressive tumours and may be an independent, unfavourable prognostic marker. However, the prognostic value of p53 in breast carcinoma is controversial [19, 28–36].

In this study, we examined the incidence of p53 in nodenegative breast carcinomas. The relationship between p53 expression and clinical and prognostic factors, including proliferation markers and receptor status, was also studied. The predictive value of p53, in relation to overall survival and disease-free survival, was calculated.

MATERIALS AND METHODS

Formalin-fixed, paraffin-embedded blocks of 174 nodenegative breast carcinomas, diagnosed between 1981 and 1986 at the Hospital Covadonga, were studied. We excluded from this group those patients who were lost to follow-up (10), and those with in situ carcinomas (11), or lobular carcinomas (2), thus limiting our study to 151 patients with infiltrating ductal carcinomas of the breast with negative axillary lymph nodes at the time of diagnosis. The patients' ages ranged from 29 to 88 years, with a mean age of 57 years. Follow-up was available in all patients with a median follow-up duration of 75 months. All patients were treated with partial, or modified radical mastectomy and axillary node dissection. Histopathological confirmation of node status was available for all patients. 41 patients received adjuvant radiotherapy. 30 patients received antioestrogen (tamoxifen) alone, and 50 radiotherapy plus tamoxifen. Chemotherapy (methotrexate/cyclophosamide/5fluorouracil) was received by 13 patients. Only 2 patients were treated with chemotherapy and tamoxifen before mastectomy. Tumour types were classified according to the WHO classification. Histological grades of tumours were scored according to Bloom and Richardson criteria.

p53 protein expression evaluation

A monoclonal mouse anti-human p53 protein DO-7 (Dako, Carpentaria, California, U.S.A.) was used that reacts with an epitope on the p53 protein, which is resistant to formalin fixation. The antibody was diluted 1:50, incubated with the slides for 1 h, at room temperature, followed by a standard avidin-biotin-peroxidase complex (ABC) technique. A microwave oven heating technique was used as the retrieval method. The tissue sections were placed in 10 mM citrate buffer at pH 6.0, and were treated in the microwave oven (750 W) for 10 min (the buffer was boiled), turning the oven on three times for 5, 2.5 and 2.5 min, with a pause of 5 and 3 min between each period. The section was kept immersed in the buffer throughout the treatment, refilling the container with distilled water if necessary. The tissue sections were left at room temperature for 20 min after the last treatment. Diaminobenzidine-hydrogen peroxidase was employed as the chromogen and a light haematoxylin counterstain was used.

Immunostaining was scored by evaluating the percentage of p53 immunopositive tumour cell nuclei. The tumours were classified into four groups as follows: negative tumours: no stained neoplastic cells; low immunoreactivity: 1–10% stained cells; moderate immunoreactivity: 10–50% stained cells and high immunoreactivity: more than 50% stained cells.

For statistical analysis, staining was evaluated according to two immunostaining patterns previously described [37]:

pattern I: high immunoreactivity (>50% cells) and pattern II: negative, low and moderate immunoreactivity.

Evaluation of proliferation markers

A recently available Ki-67 proliferating marker, MIB-1 (Immunotech, Marseille, France) was used. A microwave oven heating technique was also employed. An ABC technique, with a 1:50 antibody dilution with 1 h incubation was used. MIB-1 immunostaining results were scored, counting at least 500 cells in more than 10 high-power representative fields. Cases with less than 10% of positive cells were considered MIB-1 low labelling index (LI). Between 10% and 30% was considered as moderate MIB-1 LI and those cases with more than 30% of positive cells were considered high MIB-1 LI.

Oestrogen and progesterone receptor status evaluation

Oestrogen (Dako, Carpentaria, California, U.S.A.) (ER) and progesterone (Novocastra Laboratories, Newcastle U.K.) (PgR) receptors were evaluated immunohistochemically in formalin-fixed paraffin-embedded tumours. The microwave oven heating technique was used. A histoscore was employed for immunostaining according to staining intensity (I) and percentage (P) of stained cells: histoscore = $I \times P$. Staining intensity was evaluated as follows: negative (no staining at all); low (+) = 1; moderate (++) = 2 and strong (+ + +) = 3. Cases with less than 10 histoscore were considered as negative.

Controls

A positive control was used in all cases, usually a known positive breast tumour for p53, oestrogen and progesterone receptors and tonsil for MIB-1. Negative controls consisted of substitution of primary antibody by PBS; all other steps were followed unchanged.

Statistical analysis

Statistical analysis was performed using SPSS/WIN statistical software. The relationship between immunohistochemical staining and morphological variables was evaluated by means of a univariate chi-square test for association. Analysis of disease-free and overall survival were performed with the use of the Kaplan–Meier method. Tests for differences between curves were made with the log-rank tests. For the survival study, p53 and MIB-1 staining cases were divided into two groups: p53 (pattern I) and MIB-1 (high LI) overexpression cases and the remaining cases.

Multivariate analysis was carried out using a Cox's logistic regression, considering breast cancer deaths as events for relative risks. The following variables were included in the analysis: p53, histological and nuclear grade, hormonal receptor status, MIB-1, tumour size and tumour necrosis. Significance was established at the P < 0.05 level.

RESULTS

p53 pattern I nuclear staining (high immunoreactivity) was seen in 45 of 151 cases of breast carcinomas (30%) (Figure 1). For tumours with pattern II immunoreactivity 45 cases were totally negative (30%), 49 (32%) showed low and 12 (8%) had moderate immunoreactivity. Normal breast epithelium and stroma were negative.

Table 1 shows the correlation between tumour parameters and p53 positivity. p53 positivity was associated

1270 M. Fresno et al.

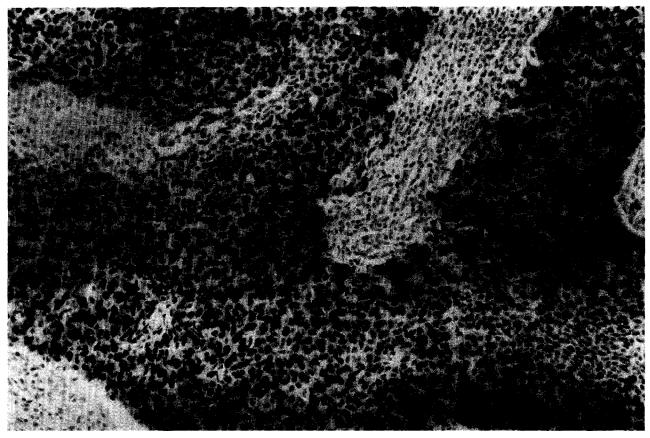


Figure 1. p53 pattern I nuclear immunoreactivity. All or nearly all cells show nuclear staining (×200).

Table 1. Correlation between p53 protein expression and biological and clinical parameters

p53 ⁺ p53 ⁻ <i>P</i> Chi							
Histological grade							
I	0	35					
II	6	44 °					
III	39	27	< 0.0001	49.49			
Nuclear grade	39	21	< 0.0001	47.47			
I I I I I I I I I I I I I I I I I I I	0	45					
_	16		< 0.0001	26.06			
II		39	< 0.0001	36.96			
III	29	22					
Receptor status*							
ER^+	9	70	< 0.0001	27.27			
ER ⁻	35	34					
PgR^+	4	36					
PgR ⁻	40	65	< 0.001	10.81			
Proliferation marker*							
Low MIB-1 LI	4	58					
Moderate MIB-1 LI	9	30	< 0.0001	49.53			
High MIB-1 LI	31	14					
Tumour size (cm)*							
Less than 1.5 cm	7	22					
Between 1.6 and 2 cm	18	54	NS	3.44			
More than 2 cm	19	29					
Necrosis							
No necrosis	6	80					
Focal	9	14	< 0.0001	66.17			
Moderate	12	11					
Extensive	18	1					

^{*}Data incomplete. ER, oestrogen receptor; PgR, progesterone receptor; LI, labelling index.

with high histological and nuclear grade (P < 0.0001) and especially with tumour necrosis (P < 0.0001); all cases with extensive tumour necrosis were pattern I stained. No association was found with tumour size. A significant association was observed between p53 positive staining and ER (P < 0.0001) and PgR negativity (P = 0.001).

The log-rank between survival curves was significant (P=0.003) with a short survival for cases with pattern I staining (Figure 2). No association was found between disease-free survival and p53 (P=0, 1) (Figure 3).

For MIB-1 evaluation (146 cases studied), every stained nucleus was considered positive, independent of intensity. A low MIB-1 LI was observed in 62 cases (45%), moderate in 39 cases (27%) and high LI in 45 cases (31%). Table 2 shows MIB-1 correlation with tumour parameters. A significant association was found between high LI and high histological and nuclear grade (P < 0.0001) as well as with tumour necrosis (P < 0.0001). No association was found with tumour size (Table 2). A significant inverse correlation was found with ER (P < 0.0001) and PgR content (P = 0.0003). The log-rank between survival curves was significant (P = 0.02) with a short survival for cases with moderate and high LI (Figure 4).

ER (148 cases studied) was positive in 53% of tumours and PgR (145 cases) in 28%. Both ER and PgR were significantly associated with histological (ER: P < 0.001; PgR: P = 0.003), nuclear grade (ER: P < 0.0001; PgR: P = 0.003) and tumour necrosis (ER: P < 0.0001; PgR: P = 0.0005).

A strong significant association was observed between p53 and MIB-1 (P < 0.0001).

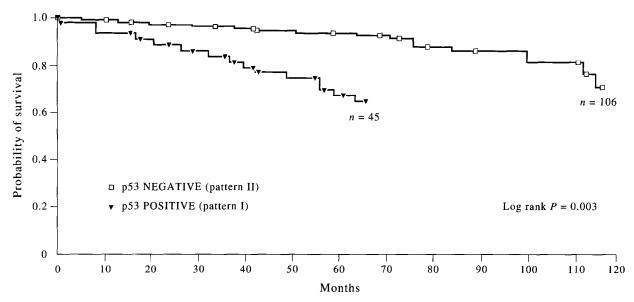


Figure 2. Overall survival of patients relative to immunohistochemical staining for p53.

A multivariate analysis was conducted to evaluate whether the correlation of p53 with survival depended on the association with the other factors indicative of a poor prognosis. The results are shown in Table 3. Overexpression of p53 protein was the strongest independent predictor of overall survival.

Cox's analysis has shown that p53 overexpression has an independent prognostic value for overall survival with a relative risk of 2.93 (95% CI 1.41–6.09).

DISCUSSION

p53 protein expression is the most widely studied of the tumour suppressor genes, present in 26-54% of primary invasive breast carcinoma [16, 28, 33, 37-39]. In intraductal tumours, the percentage has been reported to be 15-26% [15, 36]. These differences could be related to the type of monoclonal antibody [35, 38] or the immunohistochemical procedure and tissue used. Another difference among different studies is the lack of standardised immuno-

histochemical criteria to classify a tumour as p53 positive or negative. In the present study, 70% of the breast cancers were positive for p53; we found a greater number of positive tumours than other studies, probably due to the fact that we used, for formalin-fixed paraffin-embedded tissue, a microwave oven heating technique as antigen retrieval. For this reason, we used a high cut-off (50% of stained cells) in statistical analysis. Most of studies use 20–30% of stained cells as the cut-off point [19, 33, 35].

Our observations in this study of 151 node-negative breast carcinomas suggest a significant association of tumour p53 protein expression with a lack of ER and PgR reactivity and high nuclear grade. These results agree with those reported by others [16, 33, 35, 38]. A strong association between p53 immunoreactivity and necrosis has also been found by Poller and associates [15] in intraductal carcinomas. In the series of Ostrowsky and associates [28], although p53 positivity was seen in patients with a worse prognosis, no statistical significance was found. However,

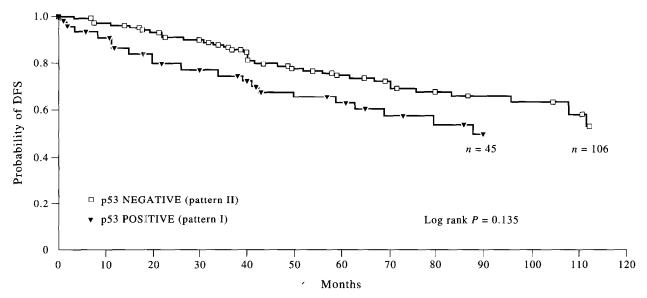


Figure 3. Disease-free survival (DFS) of patients relative to immunohistochemical staining for p53.

1272 M. Fresno et al.

Table 2. Correlation between	MIB-1	proliferation	index and	biological	and clinical	parameters

		MIB-1 LI			
	Low	Moderate	High	P	Chi
Histological grade					
I	26	5	2		
II	23	19	6		
Ш	13	15	37	< 0.0001	49.42
Nuclear grade*					
I	30	10	2		
II	24	16	13	< 0.0001	40.31
III	8	13	30		
Receptor status*					
ER ⁺	46	20	10	< 0.0001	30.63
ER-	14	19	35		
PgR^+	26	9	4		
PgR ⁻	34	29	41	< 0.001	15.71
Tumour size (cm)*					
Less than 1.5 cm	13	8	8		
Between 1.6 and 2 cm	34	19	16	NS	4.92
More than 2 cm	15	12	19		
Necrosis					
No necrosis	52	19	11		
Focal	7	11	4	< 0.0001	59.44
Moderate	2	6	15		
Extensive	1	3	15		

^{*}Data incomplete. ER, oestrogen receptor; PgR, progesterone receptor; LI, labelling index.

Barnes and associates [17] found a considerably worse prognosis in patients with p53 protein expression in the majority of their tumour cells.

The use of p53 protein overexpression may be used, according to some authors, as a diagnostic tool for malignancy, especially in cytological samples. This use may be possible only in those cases with pattern I immunostaining in which almost 100% of cells are stained, but not in cases with pattern II immunostaining because weak and focal staining has been detected in benign lesions such as fibroadenomas, benign fibrocystic disease and epithelial hyperplasia without atypia.

A significant association of MIB-1 with high histological grade, high mitotic count and an inverse relationship with ER were confirmed in our study. The strong association with p53 is in keeping with those reported by Bosari and

38associates [33], Cattoretti and co-workers [38] and Barbareschi and colleagues [16]. We also found a significant association with tumour necrosis, progesterone receptor and survival. The semiquantitative values of MIB-1 immunostaining observed in our study are higher than those reported in other studies [40], these being possibly due to the monoclonal antibody used and especially to the use of the microwave oven heating technique.

The different results obtained with p53 overexpression in breast cancer might be explained by the role of p53 protein in cancer biology. The literature on the prognostic value of p53 immunoreactivity is very controversial. Similar to our results, most studies have found a correlation between p53 expression and high tumour grade and other established indicators of poor prognosis, such as hormone receptors, high proliferation index and necrosis. Our results confirm the

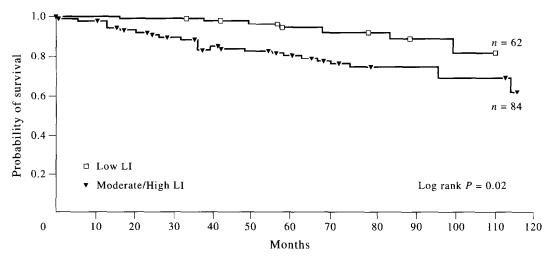


Figure 4. Overall survival of patients relative to immunohistochemical staining for MIB-1.

Table 3. p53 expression in patients with breast cancer

	Overall	survival	DFS		
Factor	Univariate P value	Multivariate P value	Univariate P value	Multivariate P value	
p53	0.003	0.004	0.135	0.463	
HG	0.207	0.245	0.062	0.408	
NG	0.008	0.154	0.493	0.862	
ER	0.066	0.598	0.042	0.041	
PgR	0.185	0.650	0.112	0.507	
MIB-1	0.01	0.182	0.23	0.618	
Size	0.066	0.128	0.151	0.110	
Necrosis	0.01	0.184	0.093	0.415	

DFS, disease-free survival; HG, histological grade; NG, nuclear grade; ER, oestrogen receptor; PgR, progesterone receptor.

independent prognostic significance of p53 overexpression reported by other investigators [19, 31, 36, 41, 42] in lymph node negative tumours and are in keeping with those studies which found an association or a trend between high p53 expression and a shorter disease-free survival [33, 34]. Although many authors have found p53 overexpression to be a useful prognostic marker, many others have only found a strong correlation with p53 immunostaining and other classical prognosticators, particularly histological grade and absence of oestrogen receptor [18, 28, 30, 35, 43].

We conclude that p53 protein expression is an additional independent biological marker in node-negative breast carcinoma. The present study shows that patients with high p53 reactivity (pattern I) have a poor prognosis. p53 expression could be used with other established prognostic factors to identify high-risk groups in node-negative breast cancer.

- Thorpe SM, Rose C, Rasmussen BB, Mouridsen HT, Bayer T, Keiding N. On the behalf of the Danish Breast Cancer Cooperative Group. Cancer Res 1987, 47, 6126-6133.
- Hollingsworth RE, Lee WH. Tumor suppressor genes: New prospects for cancer research. J Natl Cancer Inst 1991, 83, 91– 96.
- Finlay CA, Hinds PW, Levine AJ. The p53 proto-oncogene can act as a suppressor of transformation. Cell 1989, 57, 1083– 1093.
- Levine AJ, Momad J, Finley CA. The p53 tumor suppressor gene. Nature 1991, 351, 453-456.
- Nigro JM, Baker S, Preisinger A, et al. Mutations in the p53 gene occur in diverse human tumor types. Nature 1987, 342, 705-708.
- Wynford-Thomas D. Oncogenes and anti-oncogenes: The molecular basis of tumor behavior. J Pathol 1991, 165, 187–201.
- Sturzbecher HW, Maimets T, Chumakow P, et al. p53 interacts in mammalian cells: implications for cell cycle control and oncogenesis. Oncogene 1990, 5, 795–801.
- Baker SJ, Markowitz S, Fearon ER, Willson JKV, Vogelstein B. Suppression of human colorectal carcinoma cell growth by wild-type p53. Science 1990, 249, 912-915.
- Mercer WE, Shields MT, Lin D, Appella E, Ullrich SJ. Growth suppressor induced by wild-type p53 protein is accompanied by selective down-regulation of proliferating cell nuclear antigen expression. Proc Natl Acad Sci 1990, 88, 1958–1962
- Iggo R, Gatter K, Bartek J, et al. Increased expression of mutant forms of p53 oncogene in primary lung cancer. Lancet 1990, 35, 675-679.
- Lohmann D, Ruhri C, Schmitt M, Graeff H, Hofler H. Accumulation of p53 protein as an indicator for p53 gene mutation in breast cancer. Occurrence of false-positives and false-negatives. *Diagnost Mol Pathol* 1993, 2, 36-41.

- Hurlimann J, Chaubert P, Benhattar J. p53 gene alterations and p53 protein accumulation in infiltrating ductal breast carcinomas: correlation between immunohistochemical and molecular biology techniques. *Mod Pathol* 1994, 7, 423–428.
- 13. Fields S, Jang SK. Presence of a potent transcription activating sequence in the p53 protein. *Science* 1992, **249**, 1046–1051.
- 14. Lane DP, Benchimol S. p53: oncogene or anti-oncogene. Genes Devel 1990, 4, 1-8.
- Poller DN, Roberts EC, Bell JA, et al. p53 protein expression in mammary ductal carcinoma in situ. Relationship to immunohistochemical expression of oestrogen receptor and c-erbB-2 protein. Human Pathol 1993, 24, 463-468.
- 16. Barbareschi M, Leonardi E, Mauri FA, Serio G, Palma P. p53 and c-erbB-2 protein expression in breast carcinomas. An immunohistochemical study including correlations with receptor status, proliferation markers, and clinical stage in human breast cancer. Am J Clin Pathol 1992, 98, 408-418.
- Barnes DM, Dublin EA, Fisher CJ, et al. Immunohistochemical detection of p53 protein in mammary carcinoma: An important new independent indicator or prognosis? Hum Pathol 1993, 24, 469-476.
- 18. Martinazzi M, Crivelli F, Zampatti C, Martinazzi S. Relationship between p53 expression and other prognostic factors in human breast carcinoma. An immunohistochemical study. Am J Clin Pathol 1993, 100, 213-217.
- Isola J, Visakorpi T, Holli K, Kallioneimi OP. Association of overexpression of tumor suppressor protein p53 with rapid cell proliferation and poor prognosis in node-negative breast cancer patients. J Natl Cancer Inst 1992, 84, 1109-1114.
- Marks JV, Davidoff AM, Kerns BJ, et al. Overexpression and mutation of p53 in epithelial ovarian cancer. Cancer Res 1991, 51, 2979-2984.
- 21. Chang K, Ding I, Kern FG, Willinghan MC. Immuno-histochemical analysis of p53 and HER-2/neu proteins in human tumors. J Histochem Cytochem 1991, 39, 1281-1287.
- Barbareschi M, Girlando S, Mauri FA, et al. Tumor suppressor gene products, proliferation and differentiation markers expression in lung neuroendocrine neoplasms. J Pathol 1992, 166, 343-350.
- Wright C, Mellon K, Johnston P, et al. Expression of mutant p53, c-erbB-2 and the epidermal growth factor receptor in transitional cell carcinoma of the human urinary bladder. Br J Cancer 1991, 63, 967-970.
- Moch H, Sauter G, Moore D, Mihatsch MJ, Gudat F, Waldman F. p53 and erbB-2 protein overexpression are associated with early invasion and metastasis in bladder cancer. Virch Arch A Pathol Anat 1993, 423, 329-334.
- Lipponen P. Overexpression of nuclear oncoprotein in transitional cell bladder cancer and its prognostic value. Int J Cancer 1993, 53, 365-370.
- Campo E, Calle-Martin O de la, Miquel R, et al. Loss of heterozygosity of the p53 gene and p53 protein expression in human colorectal carcinomas. Cancer Res 1991, 51, 4436–4442.
- Purdie CA, O'Grady J, Piris J, et al. p53 expression in colorectal tumors. Am J Pathol 1991, 138, 807-813.
- 28. Ostrowsky JL, Sawan A, Henry C, et al. p53 expression in human breast related to survival and prognostic factors: an immunohistochemical study. *J Pathol* 1991, **164**, 75-81.
- 29. Sawan A, Randall B, Angus B, et al. Retinoblastoma and p53 gene expression related to relapse and survival in human breast cancer: an immunohistochemical study. *J Pathol* 1992, **168**, 23-28.
- Lipponen P, Aaltomaa HJ, Syrjänen S, Syrjän P. p53 protein expression in breast cancer as related to histopathological characteristics and prognosis. Int J Cancer 1993, 55, 51-56.
- Silvestrini R, Benini E, Diadone MG, Veneroni S, Boracchi P. p53 as an independent prognostic marker in lymph node-negative breast cancer patients. J Natl Cancer Inst 1993, 85, 965–970
- 32. Battifora H. p53. A word of caution. *Human Pathol* 1994, 25,
- Bosari S, Lee A, Viale G, Heatley GJ, Coggi G. Abnormal p53 immunoreactivity and prognosis in node-negative breast carcinomas with long-term follow-up. Virch Arch A Pathol Ant 1992, 421, 291-295.

1274

- Elledge RM, Clark GM, Fuqua SAW, Yu YY, Allred DC. p53
 protein accumulation detected by five different antibodies: relationship to prognosis and heat shock protein 70 in breast cancer. Cancer Res 1994, 54, 3752–3757.
- 35. Rosen PP, Lesser ML, Arroyo CD, Cranor M, Borgen P, Norton L. p53 in node-negative breast carcinoma: an immuno-histochemical study of epidemiologic risk factors, histologic features, and prognosis. *J Clin Oncol* 1995, 13, 821-830.
- Allred DC, Clark GM, Elledge R, et al. Association of p53 protein expression with tumor cell proliferation rate and clinical outcome in node-negative breast cancer. J Natl Cancer Inst 1993, 85, 200-206.
- 37. Bartek I, Bartkova I, Vojtesek B, et al. Pattern of expression of the p53 tumor suppressor gene in human breast tissue and tumors in situ and in vitro. Int J Cancer 1990, 46, 839-844.
- Cattoretti G, Rilke F, Andreola S, et al. p53 expression in breast cancer. Int J Cancer 1988, 41, 178-183.
 Umekita Y, Takasaki T, Yoshida H. Expression of p53 in
- Umekita Y, Takasaki T, Yoshida H. Expression of p53 in benign epithelial hyperplasia, atypical ductal hyperplasia, noninvasive mammary carcinoma: an immunohistochemical study. Virch Arch 1994, 424, 491–494.

- Gorczyca W, Markiewski M, Kram A, Tuziak T, Domagala W. Immunohistochemical analysis of bc1-2 and p53 expression in breast carcinomas: their correlation with Ki-67 growth fraction. Virch Arch 1995, 426, 229-233.
- Thor AD, Moore DH, Edgerton SM, et al. Accumulation of p53 tumor suppressor gene protein: an independent marker of prognosis in breast cancers. J Natl Cancer Inst 1992, 84, 845– 855
- 42. Friedrichs K, Gluba S, Eidtmann H, Jonat W. Overexpression of p53 and prognosis in breast cancer. *Cancer* 1993, 72, 3641-3647
- 43. Davidoff AM, Herndon JE, Glover NS, et al. Relation between p53 overexpression and established prognostic factors in breast cancer. Surgery 1991, 110, 259–264.

Acknowledgements—The authors thank Aurora Fernández-García and Inés Arguelles from the Section of Immunohistochemistry for immunohistochemical staining.